The use of mass spectrometry in lipidomics

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Outlines

- **Brief introduction to lipidomics**
- **Analytical methodology: MS/MS structure elucidation of phospholipids**
- **Phospholipid analysis in lean and ob/ob mice by mass spectrometry**
- **LC-MS/MS quantification of ceramides**

Lipidomics- A comprehensive analysis of lipid molecules in response to cellular stress and challenges

Lipids are very important!!

Nutrition

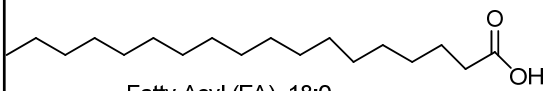
-Energy source

-Energy storage

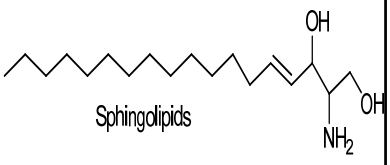
Nutrition related diseases-
Atherosclerosis, diabetes

Phospholipids are essential- membrane composition/ functional state of cells

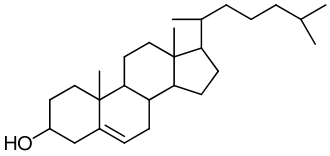
Structures of different lipids classes



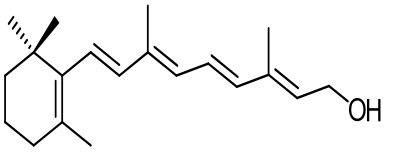
Fatty Acyl (FA), 18:0
saturated/unsaturated



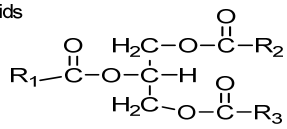
Sphingolipids



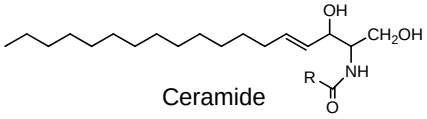
Sterol lipids



Prenol lipids (retinol)

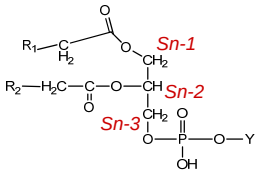


Glycerolipid, R =
saturated/unsaturated FA

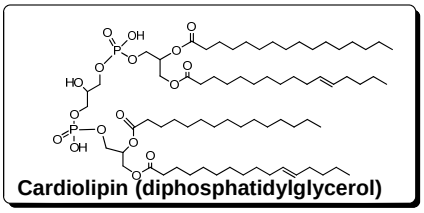


Ceramide

Structures of main phospholipids



Phosphatidylethanolamine (PE)
Y= NH_2
Phosphatidylcholine (PC)
Y= $N(CH_3)_3$
Phosphatidylglycerol (PG)
Y= OH OH
Phosphatidylserine (PS)
Y= O OH NH_2
Phosphatidylinositol (PI)
Y= HO HO OH OH OH



Cardiolipin (diphosphatidylglycerol)

Extraction of lipids by Bligh/Dyer method

- To a homogenized sample (1 ml containing internal standards) add methanol (2.5 ml) and chloroform (1.25 ml), sonicate by 4-5 bursts and added 1.0 ml water and 1.25 ml chloroform additionally and vigorously shaken.
- Centrifuge (1,000 x g) for 2 min and separate the chloroform layer (bottom layer) and repeat the process twice.
- Combine the chloroform soluble phase and evaporate to dryness and stored at -20 ^{o}C untill analysis.

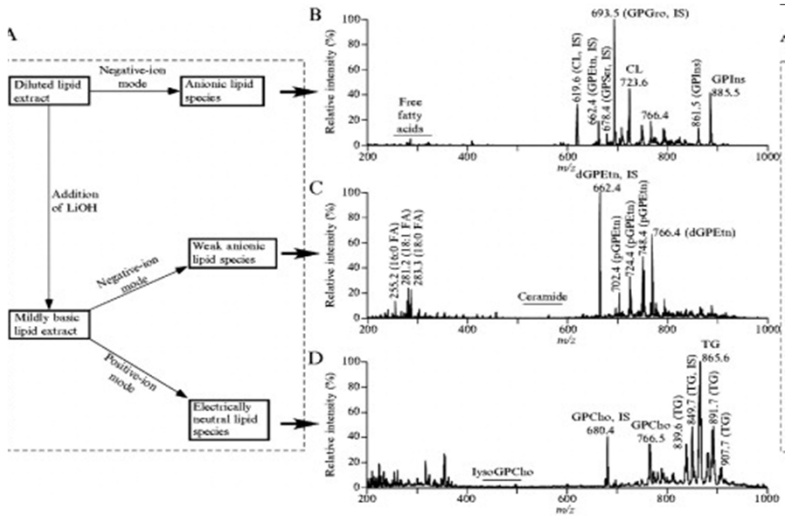
Shotgun lipidomics: intrasource separation of lipids for quantitative lipidomics

Group	Electrical Propensity	Lipid Classes
Anionic lipids	Carry net negative charge(s) at physiological pH	Cardiolipin, acylCoA, sulfatide, PtdIns (PtdInsP, PtdInsP$_2$, PtdInsP$_3$), PtdGro, PtdSer, PtdH, etc.
Weak anionic lipids	Carry a net negative charge at alkaline pH	PE, lysoPE, ceramide, NEFA, eicosanoids, etc.
Neutral polar lipids	Neutral at alkaline pH	PC, lysoPC, SM, glycolipid, TAG, etc.
Special lipids	Vary	Acylcarnitine, sterols, etc.

The ionization efficiency of an analyte greatly depends on the electrical propensity of an individual analyte in its own microenvironment to lose or gain a charge

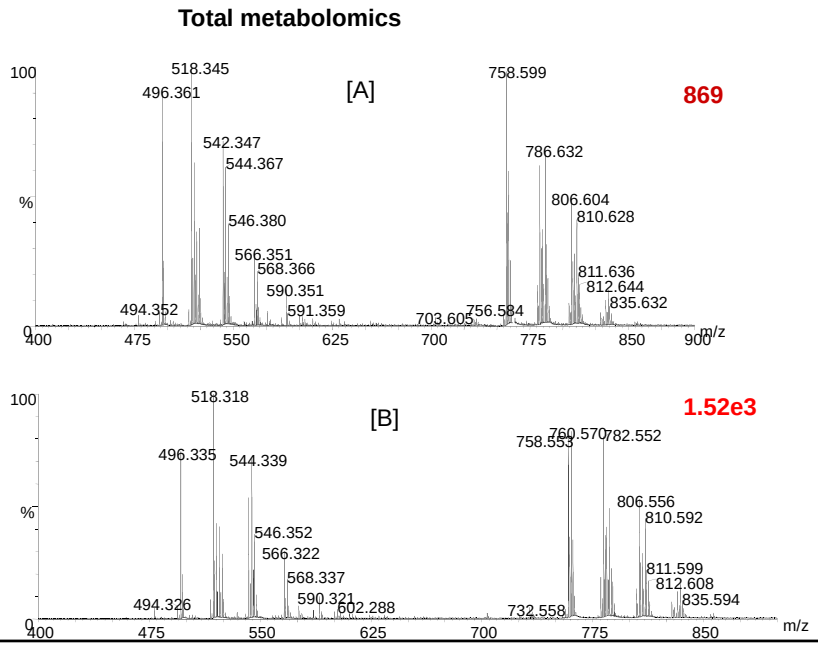
Source: Gross and Han,, 2004

Application of shotgun lipidomics: intra-source separation of lipids

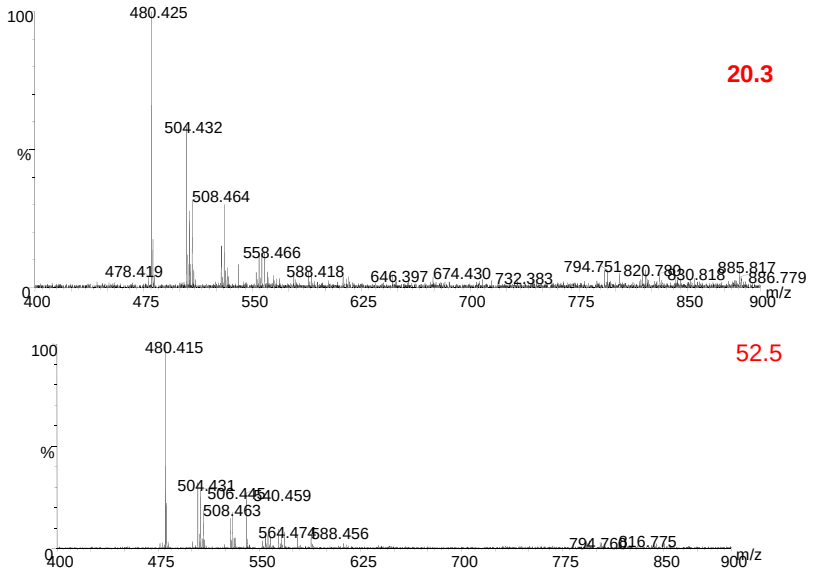


Source: Gross and Han, methods in Enzymology, 2007

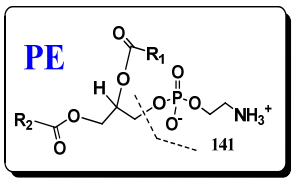
Total scan of metabolites (Q1 SCAN + ion mode) for a plasma sample obtained from lean mouse [A]; ob/ob mouse



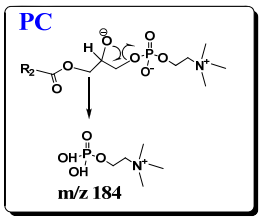
Total scan of metabolites (Q1 SCAN -ve ion mode) for a plasma sample obtained from lean mouse [A]; ob/ob mouse

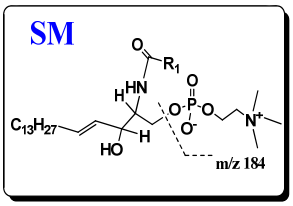


Profiling phospholipids and sphingosines in a complex mixture using MS/MS

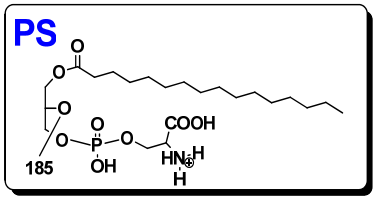


PE
Neutral Loss scan 141



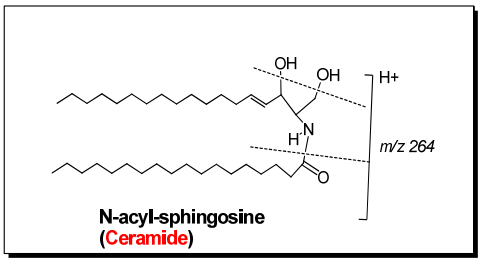


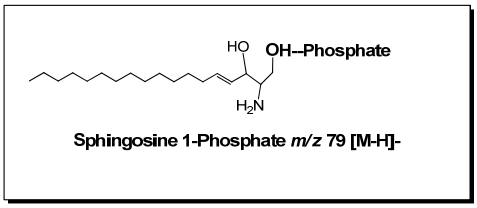
PC & SM
Precursor ion scan 184



PS
Neutral Loss scan 185

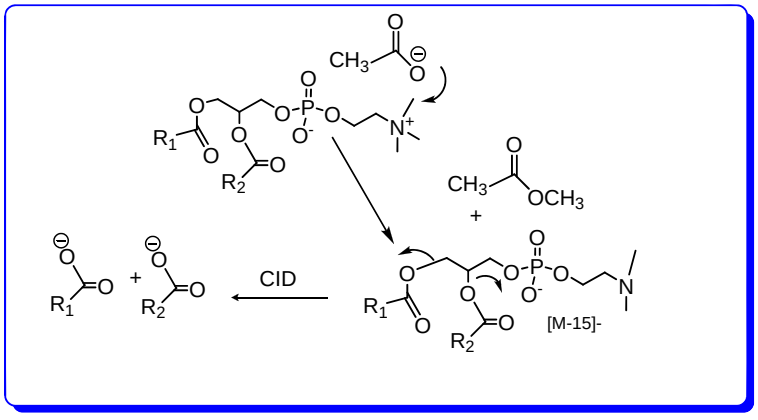
How to profile sphingolipids in a complex mixture using MS/MS?





***m/z* 264 is a characteristic ion for all compounds containing a sphingosine backbone**

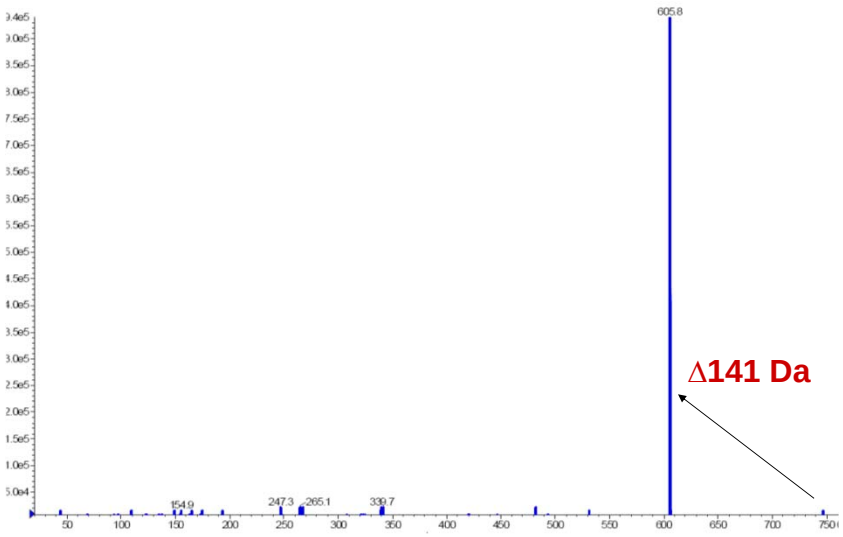
Phosphatidylcholine loses a methyl group to form a negatively charged, pseudomolecular ion



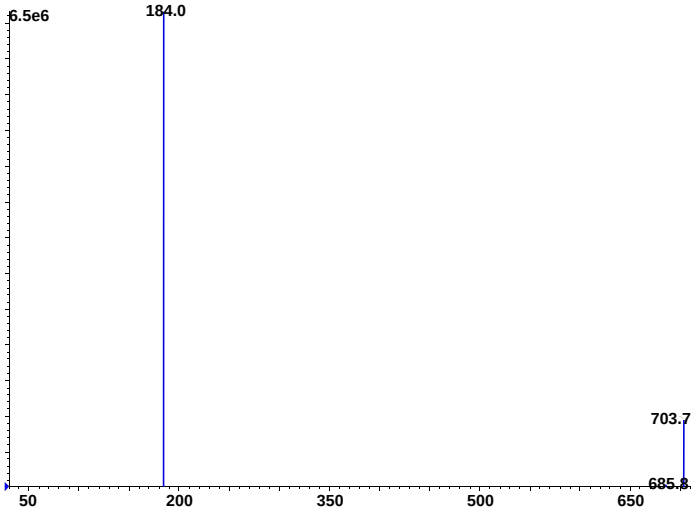
Phospholipids may undergo demethylation and then the loss of the fatty acyl groups from glycerophosphocholine backbone.

MS/MS of m/z 746.90: PE Std

Neutral loss of 141 is a characteristic for detecting PE



MS/MS of sphingomyelin standard (2S,3R,4E)-2-acylaminooctadec-4-ene-3-hydroxy-1-Phosphocholine m/z 703.7

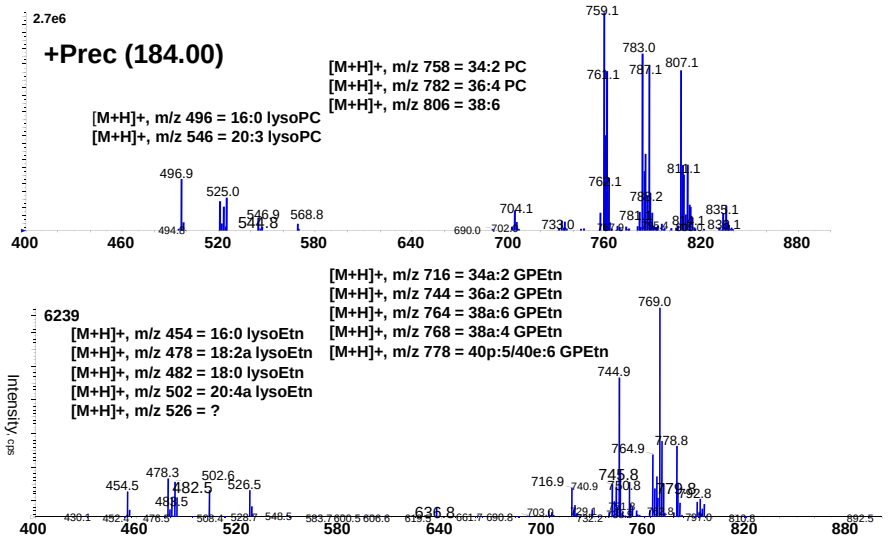


ESI-MS/MS analyses of various lipids

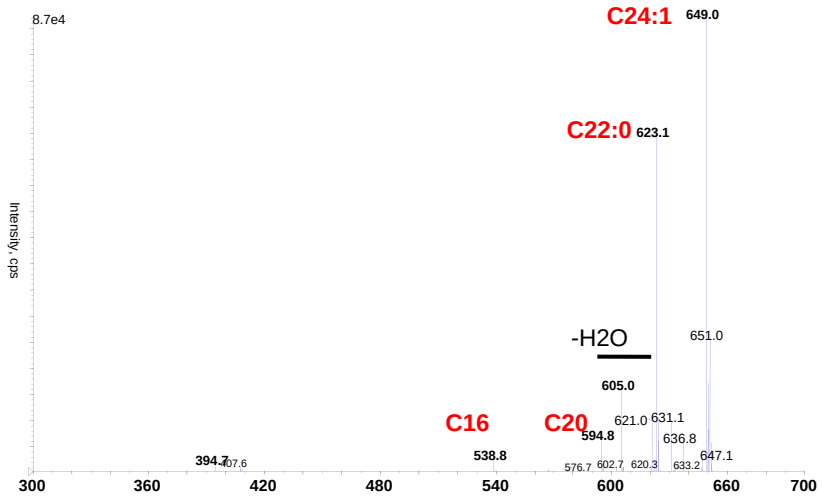
Lipid Class(s)	Precursor Ion	MS/MS Mode & Conditions	Fragment
cardiolipin	$[M-2H]^{2-}$	PI, *m/z* 153.0, 35 eV	glycerol phosphate derivative
PtdGro, PtdH	$[M-H]^-$	PI, *m/z* 153.0, 35 eV, *	glycerol phosphate derivative
PtdIns	$[M-H]^-$	PI, *m/z* 241.1, 45 eV	cyclic Inositol phosphate
		PI, *m/z* 153.0, 35 eV	glycerol phosphate derivative
PtdInsP	$[M-H]^-$	PI, *m/z* 321.1, 53 eV	phosphoinositol phosphate
$PtdInsP_2$	$[M-H]^-$	PI, *m/z* 401.1, 62 eV	diphosphoinositol phosphate
PtdSer	$[M-H]^-$	NL, 87.0 amu, 25 eV, *	serine
		PI, *m/z* 153.0, 35 eV	glycerol phosphate derivative
sulfatide	$[M-H]^-$	PI, *m/z* 97.0, 65 eV	sulfate
acylCoA	$[M-2H]^{2-}$	PI, *m/z* 339.0, 30 eV, *	doubly-charged CoA derivative
PE, lysoPE	$[M-H]^-$	PI, *m/z* 196.0, 50 eV	glycerol phosphoethanolamine derivative
ceramide	$[M-H]^-$	NL, 256.2 amu, 32 eV *	
		NL, 327.3 amu, 32 eV	
		NL, 240.2 amu, 32 eV *	2-*trans*-palmitoyl alcohol
PC, lysoPC, SM	$[M+Li(Na)]^+$	NL, 59.1 amu, -28 eV, *	trimethylamine
	$[M+Li(Na)]^+$	NL, 183.1 amu, -32 eV	phosphocholine
	$[M+Li]^+$	NL, 189.1 amu, -42 eV	lithium cholinephosphate
	$[M+Na]^+$	NL, 205.1 amu, -35 eV	sodium cholinephosphate
	$[M+H]^+$	PI, *m/z* 184.1, -30 eV, *	phosphocholine
	$[M+Cl]^-$	NL, 50.0 amu, 24 eV, *	methylchloride
cerebroside	$[M+Li]^+$	NL, 162.2, -50 eV, *	
	$[M+Cl]^-$	NL, 36.0 amu, 30 eV	hydrogen chloride
MGDG	$[M+Li(Na)]^+$	PI, *m/z* 227(243), -45 eV	Li(Na)+galactose derivative
DGDG	$[M+Li(Na)]^+$	PI, *m/z* 227(243), -66 eV	Li(Na)+galactose derivative
acylcarnitine	$[M+H]^+$	PI, *m/z* 85.1, -20 eV, *	carnitine
chol. ester	$[M+NH_4]^+$	PI, *m/z* 369.3, -50 eV, *	cholestane cation
TAG	$[M+Li]^+$	NL, X amu, -35 eV	a fatty acid

Source: Gross and Han,, 2004

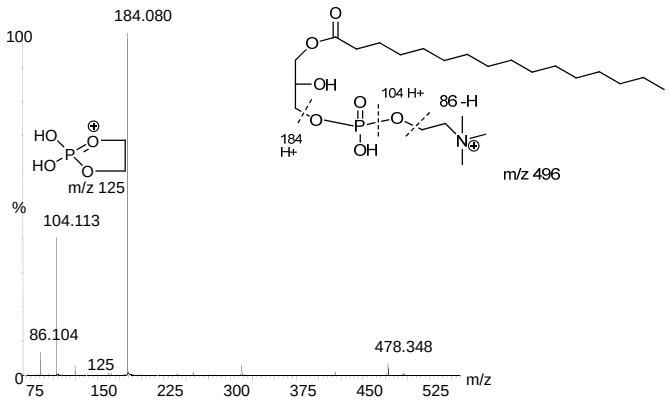
Profiling of phospholipids using precursor ion m/z 184 and neutral loss scan 141 for PC, SM and PE



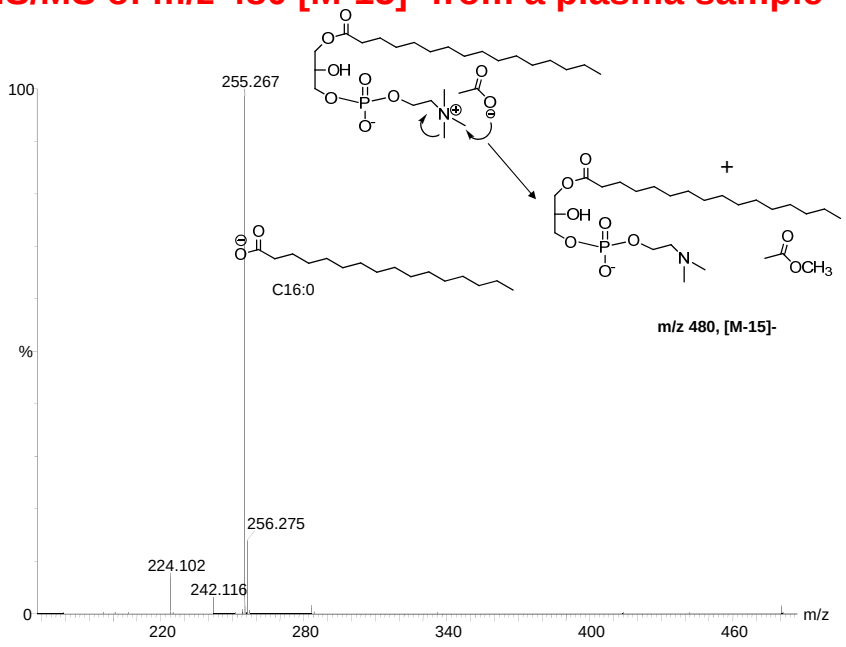
Precursor ion scan m/z 264 in +ve ion mode is specific to identify ceramides in a sample



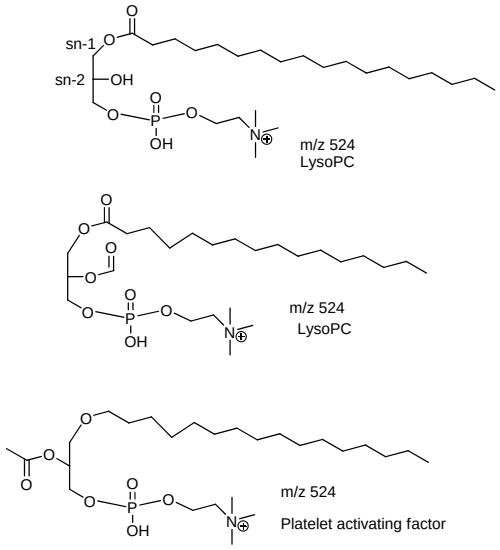
MSMS fragmentation of *m/z* 496 obtained from a plasma sample



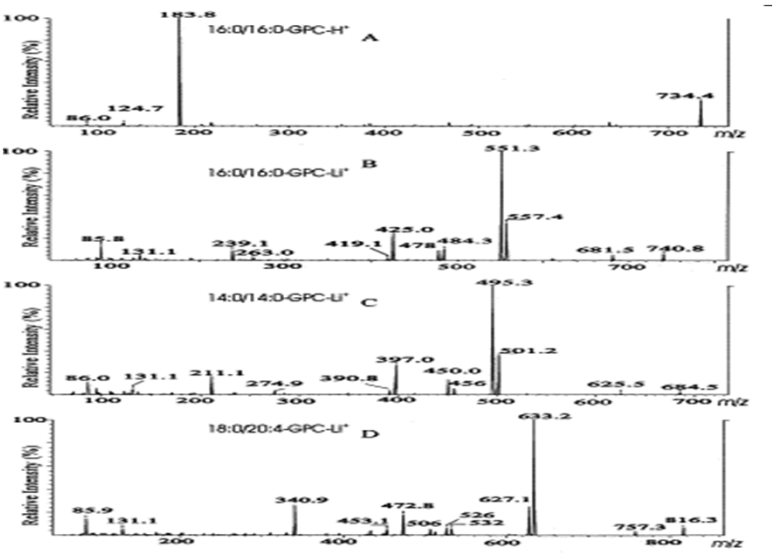
MS/MS of m/z 480 [M-15]- from a plasma sample



Several isomeric compounds exits and unambiguous identification is a challenge

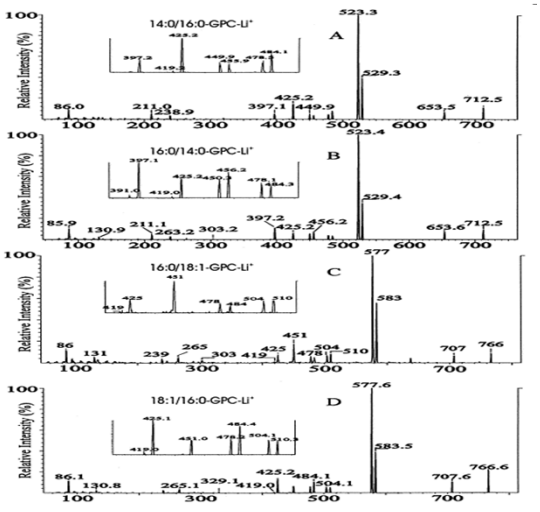


Lithiated adducts of phosphocholine provide more structural information in their MS/MS spectra



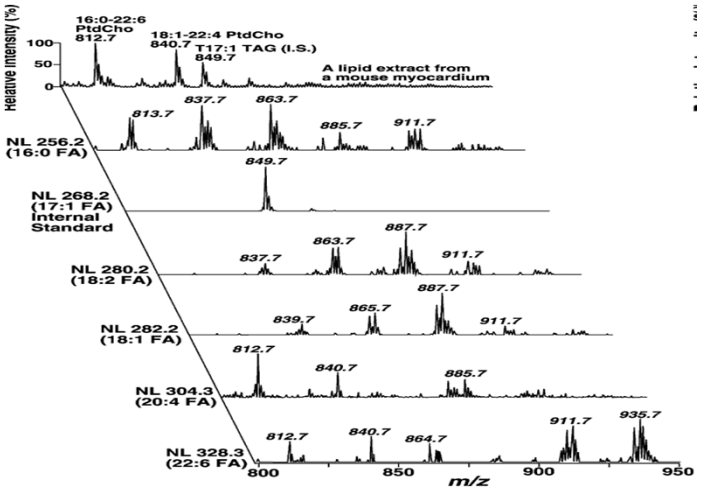
Source: Hsu et al. J. Am Soc. Mass Spectrom, 1998

Relative abundances of product ion can be used to distinguish positional isomers of lithiated phospholipids



Source: Hsu et al. J. Am Soc. Mass Spectrom, 1998

Neutral loss scans can be used to profile triacylglycerides (TAG)



Source: Han and Gross, 2004

Library search for eicosanoid http://www.lipidmaps.org/

LIPID MAPS -- LIPID Metabolites And Pathways Strategy

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LIPID Metabolites And Pathways Strategy

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LMSD: Lipid classification search results

Fatty Acyls [FA] (W) --> Eicosanoids [FA03]

LM_ID	Common Name	Systematic Name	Formula	Mass
LMFA03000001	8(9)-EpETE	(+/-)-8(9)-epoxy-5Z,11Z,14Z,17Z-eicosatetraenoic acid	$C_{20}H_{30}O_3$	318.22
LMFA03000002	11(12)-EpETE	(+/-)-11(12)-epoxy-5Z,8Z,14Z,17Z-eicosatetraenoic acid	$C_{20}H_{30}O_3$	318.22
LMFA03000003	14(15)-EpETE	(+/-)-14(15)-epoxy-5Z,8Z,11Z,17Z-eicosatetraenoic acid	$C_{20}H_{30}O_3$	318.22
LMFA03000004	17(18)-EpETE	(+/-)-17(18)-epoxy-5Z,8Z,11Z,14Z-eicosatetraenoic acid	$C_{20}H_{30}O_3$	318.22
LMFA03000005	11(R)-HEDE	11R-hydroxy-12E,14Z-eicosadienoic acid	$C_{20}H_{36}O_3$	324.27
LMFA03000006	17R,18S-EpETE	17R,18S-epoxy-5Z,8Z,11Z,14Z-eicosatetraenoic acid	$C_{20}H_{30}O_3$	318.22
LMFA03000008	15(R)-HEDE	15R-hydroxy-11Z-13E-eicosadienoic acid	$C_{20}H_{36}O_3$	324.27
LMFA03000009	11S-HEDE	11S-hydroxy-12E,14Z-eicosadienoic acid	$C_{20}H_{36}O_3$	324.27
LMFA03010000	Prostanoic acid skeleton	-	-	-
LMFA03010001	6-keto-PGF1α	6-oxo-9S,11R,15S-trihydroxy-13E-prostenoic acid	$C_{20}H_{34}O_6$	370.24
LMFA03010002	PGF2α	9S,11R,15S-trihydroxy-5Z,13E-prostadienoic acid	$C_{20}H_{34}O_5$	354.24
LMFA03010003	PGE2 (W)	9-oxo-11R,15S-dihydroxy-5Z,13E-prostadienoic acid	$C_{20}H_{32}O_5$	352.22
LMFA03010004	PGD2 (W)	9S,15S-dihydroxy-11-oxo-5Z,13E-prostadienoic acid	$C_{20}H_{32}O_5$	352.22
LMFA03010005	PGA1	9-oxo-15S-hydroxy-10Z,13E-prostadienoic acid	$C_{20}H_{32}O_4$	336.23
LMFA03010006	PGF2α-d4	9S,11R,15S-trihydroxy-5Z,13E-prostadienoic acid (3,3,4,4-d4)	$C_{20}H_{30}D_4O_5$	358.27
LMFA03010007	PGD2-d4	9S,15S-dihydroxy-11-oxo-5Z,13E-prostadienoic acid (3,3,4,4-d4)	$C_{20}H_{28}D_4O_5$	356.25
LMFA03010008	PGE2-d4	11R,15S-dihydroxy-9-oxo-5Z,13E-prostadienoic acid (3,3,4,4-d4)	$C_{20}H_{28}D_4O_5$	356.25
LMFA03010009	PGG2	9S,11R-epidioxy-15S-hydroperoxy-5Z,13E-	$C_{20}H_{32}O_6$	368.22

LIPID Metabolites And Pathways Strategy

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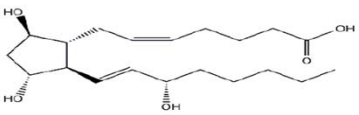
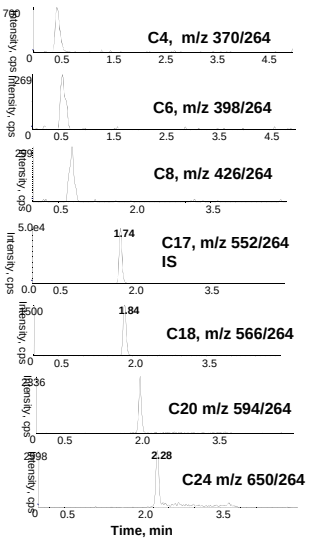
Structure database (LMSD)

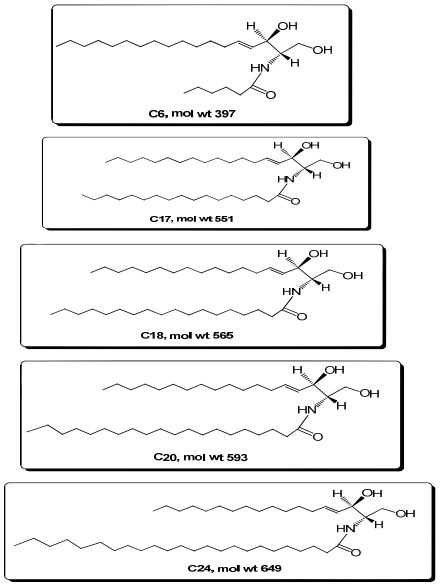
LMFA03010025



LM ID	LMFA03010025
Common Name	PGF2β
Systematic Name	9R,11R,15S-trihydroxy-5Z,13E-prostadienoic acid
Synonyms	-
Exact Mass	354.24
Formula	$C_{20}H_{34}O_5$
Category	Fatty Acyls [FA]
Main Class	Eicosanoids [FA03]
Sub Class	Prostaglandins [FA0301]
LIPIDBANK ID	XPR1764
PubChem Substance ID (SID)	4265968
KEGG ID	-

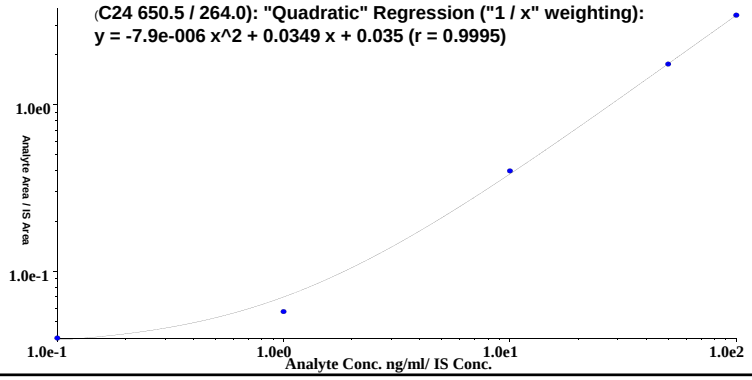
MRM chromatograms showing simultaneous determination of ceramides (C4-C24)



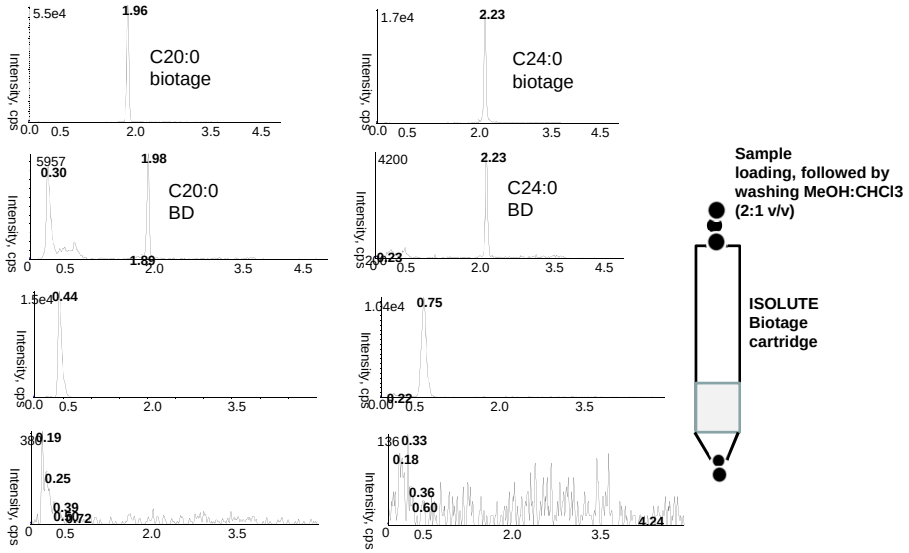


A linear response for Cer C24:0 was observed over a range of 0.1-100 ng/ml with correlation coefficient greater than 0.99

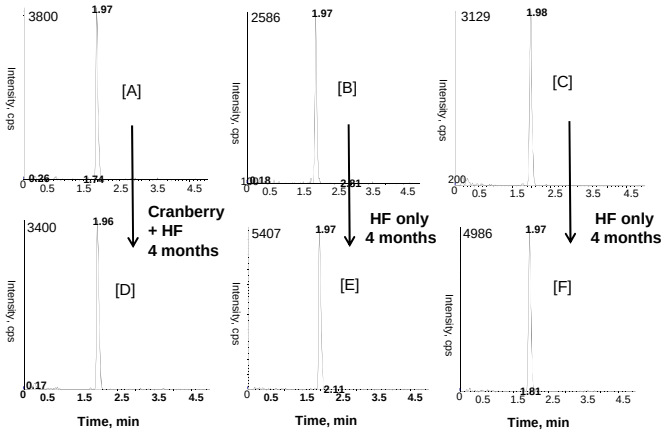
Sample Name	Analyte Peak Name	Calculated Concentration (ng/mL)	Accuracy (%)
Ceramide Standard 100 ng/ml	C24 650.5 / 264.0	100	100
Ceramide Standard 50 ng/ml	C24 650.5 / 264.0	49.8	99.6
Ceramide Standard 10 ng/ml	C24 650.5 / 264.0	10.5	105
Ceramide Standard 1 ng/ml	C24 650.5 / 264.0	0.634	63.4
Ceramide Standard 0.1 ng/ml	C24 650.5 / 264.0	0.132	132



Sample preparation of ceramides in plasma; Poor recoveries of non-polar ceramides Biotage (supported liquid extraction), but better for polar ceramides



Cranberry fruit powder treatment reduced the HF induced increased levels of Ceramide C20 in rats



[A]-[C] represent base line plasma ceramide C20 (594/264) from three animals
[D] after 4 months treatment with cranberry (1 g/kg b. w. and high fat diet
[E] & [F] after 4 months treatment with high fat diet only

Conclusions

- **Shotgun lipidomics approaches are high throughput and applicable to perform profiling as well as quantitative analysis of various lipids in biological samples.**
- **Tandem mass spectrometry analysis of phospholipids in +ve ion mode characterizes phospholipid polar head groups, whereas –ve ion mode provide fatty acid chain structural information**
- **Identification of phospholipids at a molecular level present a great challenge due to their structural diversity and dynamic metabolism.**
- **A rapid five minute liquid chromatography tandem mass spectrometry (LC-MS/MS) method operating in multiple reaction ion monitoring mode (MRM) was developed for identification and simultaneous quantification of six ceramides.**